

## HEMATOLOGIC DISEASE AT OLDER AGE

# Aging of hematopoietic stem cells

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**Hematopoietic stem cells (HSCs) ensure a balanced production of all blood cells throughout life. As they age, HSCs gradually lose their self-renewal and regenerative potential, whereas the occurrence of cellular derailment strongly increases. Here we review our current understanding of the molecular mechanisms that contribute to HSC aging. We argue that most of the causes that**

**underlie HSC aging result from cell-intrinsic pathways, and reflect on which aspects of the aging process may be reversible. Because many hematological pathologies are strongly age-associated, strategies to intervene in aspects of the stem cell aging process may have significant clinical relevance. (*Blood*. 2018;131(5):479-487)**

## Introduction

The average human life expectancy has increased consistently during the past 150 years. The absolute number of elderly people has increased substantially in many societies, and this is not likely to come to an end soon. Although it is evident that many elderly people reach advanced ages in healthy conditions, the prevalence of age-dependent disease and the average age of patients who enter the clinic are increasing. This is also true for patients that suffer from hematological diseases because many hematological conditions are strongly age-dependent. From a clinical perspective, this raises 2 related, yet distinct, considerations. First, how does the hematopoietic system change with age, how does this lead to a spectrum of hematological conditions, and would it be possible to pharmacologically intervene in this process? Second, if hematological disease presents in an elderly patient, should this be treated differently than if it had occurred in a young patient? Although the latter is also of significant clinical interest,<sup>1</sup> this manuscript will focus on our understanding of the molecular mechanisms that make hematopoietic stem cells (HSCs) age. Only if we understand how HSC age, we can begin exploring opportunities to prevent, delay, or even reverse aspects of the aging process.

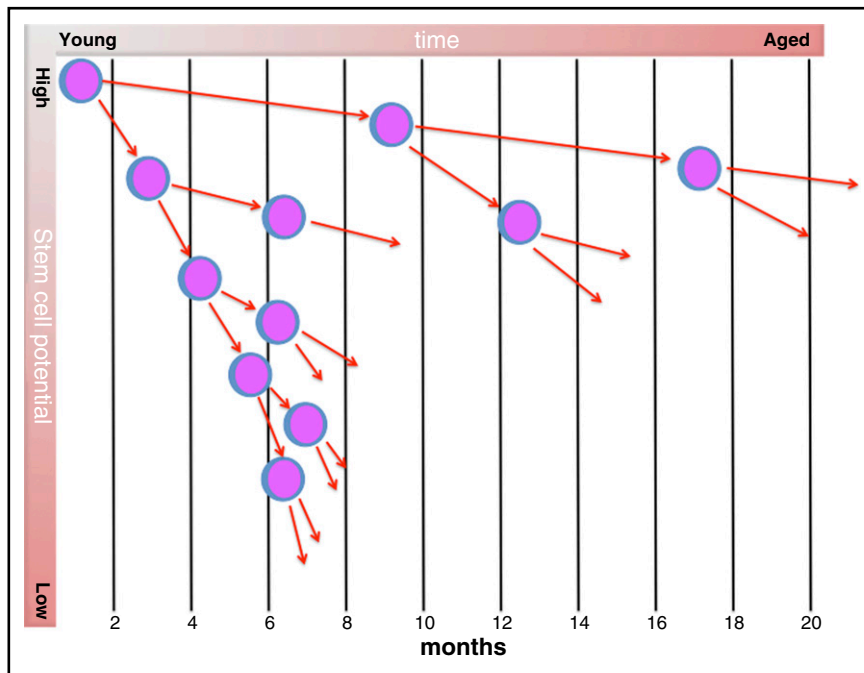
## HSC self-renewal and aging

Although research on aging has for a long time been relatively descriptive, much progress has been made in the past decade to uncover the molecular drivers of biological aging. An influential review paper that describes the hallmarks of aging provides a comprehensive overview of the various pathways that are believed to result in age-dependent cellular and organismal functional decline.<sup>2</sup> One of these so-called hallmarks relates to stem cell exhaustion. Stem cell exhaustion refers to the gradual functional decline of adult, tissue-specific stem cells to maintain homeostasis of the tissue in which they reside. Stem cells are critically defined by their ability to self-renew, a concept that seems difficult to reconcile with the notion of aging; either stem cells are able to self-renew and therefore not age, or

alternatively, stem cells age and therefore do not truly self-renew.

In the hematopoietic system, there are multiple indications that stem cells do not formally self-renew, or at least have a restricted self-renewal potential and are therefore fundamentally different from pluripotent embryonic stem or pluripotent stem cells. What are these indications? Historically, experimental hematologists have carried out serial stem cell transplantations in mice to assess the potential of these cells to multiply in vivo in myeloablatively conditioned recipients.<sup>3</sup> Although serial transplantations are obviously quite artificial (yet, they have also been performed in rare patients),<sup>4</sup> collectively these studies document that after transplant the HSC compartment never returns to normal values.<sup>5-7</sup> If stem cells are submitted to increased proliferative stress (by transplanting a low number, or even a single stem cell<sup>8</sup> or by initiating serial transplantations in short intervals),<sup>9</sup> self-renewal is impeded more severely. Most notably, if stem cells that were first harvested from an aged donor mouse are serially transplanted in young recipients, the self-renewal capacity is much lower compared with stem cells isolated from young donor mice.<sup>10</sup> If stem cells from a short-lived mouse strain are compared with stem cells from a long-lived mouse strain, the latter stem cells outcompete their shorter lived counterparts.<sup>11</sup> In general, in all studies in which young HSC were competed in transplantation studies against aged stem cells, without exception the young stem cells are functionally superior.<sup>12-16</sup> These young stem cells produce, at the single-cell level, more mature peripheral blood cells<sup>10,14</sup> and are better able to produce both myeloid and lymphoid cells in a balanced manner.<sup>10,17</sup>

There is ample evidence that during steady-state conditions the most primitive HSC are quiescent and only divide rarely. The concept of HSC quiescence emerges from studies that have shown that stem cells are refractory to cell cycle-specific cytotoxic drugs, such as 5-fluorouracil,<sup>18</sup> that stem cells are not easily labeled with 5-bromo-2'-deoxyuridine,<sup>19,20</sup> and that it can take



**Figure 1. Hypothetical tracing of the offspring of a single HSC during aging.** In mice, the most primitive HSC are believed to cycle only once every ~4 months.<sup>23</sup> With each cell division, daughter cells lose developmental (long-term repopulating) potential, such that each daughter is less potent than its ancestor. Cell-cycle times decrease with developmental stage. In young mice, the pool of stem cells is small, but the potency of each stem cell is high. In aged mice, the pool of stem cells has expanded, but their functionality is restricted. Adapted from Van Zant et al.<sup>24</sup> and Jung et al.<sup>25</sup>

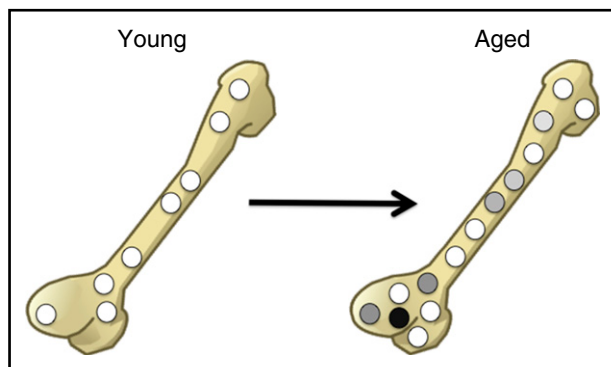
many months before they contribute to hematopoiesis after transplant.<sup>21,22</sup> Very recently, an elegant study showed that the most primitive of all HSC may undergo only 4 to 5 divisions in the lifetime of a mouse.<sup>23</sup> This study suggested that HSC possess some sort of cellular memory and that the typical age-dependent phenotype of HSC only emerges after HSCs have divided 5 times. A very similar concept had been hypothesized to exist by 1 of us more than 2 decades ago.<sup>24</sup> Collectively, this indicates that most of the proliferative burden resides with committed progenitors, and that during steady-state hematopoiesis the most primitive stem cells are largely inactive. It seems plausible that with each cell division, the potential of a HSC to contribute to blood cell production declines and that, simultaneously, the pool of stem cells with reduced potential increases to compensate for loss of function of individual stem cells (Figure 1).

Most of what we know about HSC aging is based on studies that have used the mouse as an experimental model. As a consequence, it is not fully clear to what extent these molecular mechanisms also play a role in human HSC aging. However, decades of research have shown that, conceptually, the formation of blood cells in mouse and human is identical, and it is plausible that mechanisms that contribute to stem cell aging in mice also do so in human. Indeed, it has been demonstrated that telomeres progressively shorten in human HSCs isolated from fetal liver, cord blood, or adult bone marrow, which is associated with a strongly reduced proliferative potential.<sup>26</sup> In addition, an age-dependent increase in the frequency of putative stem cells, coinciding with impaired functionality and reduced lymphoid potential, has also been in the human system.<sup>27-29</sup> However, it should be recognized that essentially all experimental studies have used C57BL/6 mice and that aging (also of the hematopoietic system) is at least qualitatively different in distinct strains of mice.<sup>30,31</sup> These genetic disparities are also likely to be prominent in human.

## Heterogeneity of HSC aging

Although there is consensus on the functional decline of aged HSC in the mouse, the molecular mechanisms that contribute to such stem cell aging are less clear and are at times disputed. Our limited insight into the molecular mechanisms that cause HSC to age may result from the fact that most studies in this field have been performed using populations of inevitably heterogeneous HSCs. Although flow cytometry has allowed the prospective isolation of single cells, the purity of HSCs, at least when assayed in transplantation experiments, is never more than 50%. Therefore, even after the most stringent enrichment protocols, many non-HSCs (mostly progenitors) remain. This is particularly problematic in the aging field because there is solid evidence that the functional heterogeneity of the HSC population increases with age. Although many HSCs in young mice behave qualitatively similarly, in aged mice that display an overall increase in HSC pool size, individual HSCs behave very differently.<sup>10,13,17,32,33</sup> Therefore, it remains possible that even in aged mice, there are still substantial numbers of very potent ("young-like") HSCs, which are diluted by an expanded pool of less potent aged cells (Figure 2).

To provide further insight into the age-dependent increase in heterogeneity of the HSC pool, the identification of cell surface markers that allow prospectively isolating of these different populations will be required. Multiple single-cell RNA sequencing studies have been performed to elucidate at the molecular level how heterogeneity may be explained.<sup>34-36</sup> Combinatorial single-cell techniques such as single-cell transplants, flow cytometry, and single-cell RNA have already begun to and will continue to identify HSC subsets that contribute to aging phenotypes. This strategy will boost research into molecular mechanisms of aging through the ability of identifying candidate marker genes (eg, receptors) for the isolation and functional characterization of "aged" HSCs.



**Figure 2.** In aged mice, the absolute number of cells with regenerative potential increases, but the extent to which individual aged cells contribute to blood cell production becomes highly variable. The relative frequency of stem cells with high regenerative potential (white) compared with cells with low regenerative potential (gray and black) thus decreases upon aging.

## Cell-intrinsic vs cell-extrinsic mechanisms

A topic of much dispute is whether HSC aging is caused by cell-intrinsic or cell-extrinsic parameters. This is not only of academic importance, but also has major implications for potential future interventions. If stem cell aging were largely intrinsically controlled, this would render putative interventions more cumbersome than if aging were the result of an extrinsic perturbation. Experiments with parabiotic mice have attracted attention (understandably, also from the lay press) because the suggestion has been raised that unidentified humoral factors that circulate in the blood of young mice could possibly restore cellular functioning in aged mice. Several studies have reported beneficial, arguably antiaging, effects of young blood on aged vasculature, brain, and muscle,<sup>37-41</sup> but no beneficial effects on aged HSC have been reported. It is important here to remember that essentially all we know on the phenotype of aged HSC is based on studies in which old stem cells were transplanted into lethally irradiated young recipients.<sup>5,10-12,15</sup> That aged HSC are functionally impaired compared with their young counterparts upon transplantation in young mice strongly suggests that HSC aging manifests largely as a consequence of cell-intrinsic molecular changes. Thus, although cell-intrinsic changes may be partially dependent on and initiated by changes that occur in the bone marrow microenvironment, transplanting aged HSCs in a young environment apparently cannot reverse these changes. It is evident that during aging many cellular and structural changes occur in the bone marrow microenvironment,<sup>42</sup> and it is possible that these as-yet poorly understood cell extrinsic changes affect stem cell intrinsically. Indeed, it has been demonstrated that young bone marrow cells, when transplanted into aged recipients, engraft worse than when transplanted into young recipients.<sup>12,43</sup> Also, hematological malignancies can result from a defective bone marrow environment.<sup>44</sup>

## Cell-intrinsic mechanisms that cause HSC aging

Multiple molecular and cell-intrinsic mechanisms have been reported to contribute to the age-associated decline of HSC functioning (partially reviewed in Geiger et al<sup>45</sup>). Although

mechanistically it may be feasible, and even useful, to separately discuss these multiple aging pathways, in effect they are likely to be highly interdependent and interconnected. Although some of these cell-intrinsic causes of aging are unlikely to be reversible, several others might allow interventions and thus could potentially be explored for pharmacological targeting (Figure 3).

### DNA damage

The most irreversible cause of HSC aging relates to the accumulation of random DNA damage. Mice, or patients for that matter, suffering from mutations in genes encoding for proteins involved in DNA repair display many aspects of premature stem cell aging.<sup>46-48</sup> In addition, aged HSC accumulate signatures of widespread DNA damage, including  $\gamma$ -H2AX foci.<sup>49</sup> To what extent normal HSC aging is caused by genetic damage remains unclear. Conceptually, if indeed HSC are largely quiescent and divide only rarely in the lifetime of a mouse, it is difficult to understand how accumulation of DNA damage could causally contribute to stem cell dysfunctioning. However, it is possible that cells immediately downstream of quiescent HSCs, which have been shown to comprise mostly myeloid-biased HSCs,<sup>50</sup> are target cells to accumulate DNA damage.<sup>51</sup>

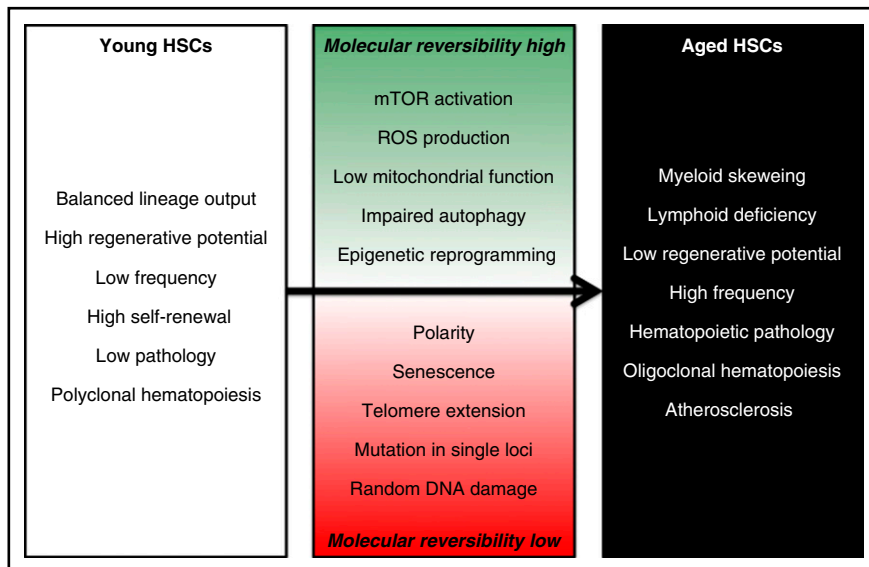
Beyond random DNA damage, it has recently been shown that in healthy elderly individuals, DNA mutations in specific loci are associated with the establishment of clonal hematopoiesis<sup>52</sup> (discussed later). A specific kind of DNA damage is caused by erosion of telomeres. Although the involvement of telomere shortening in the functional decline of HSC is particularly evident in human,<sup>53</sup> in mice with long telomeres, the inability to maintain telomere length is associated with severe HSC malfunctioning.<sup>54</sup> Although the length of telomeres in HSC can be increased by enforced overexpression of telomerase, in mice this does not rescue functional impairment.<sup>55</sup>

### Senescence

In many tissues, irreversibly cell cycle-arrested senescent cells accumulate during normal aging.<sup>56</sup> Conceptually, senescence of stem cells is a “contradictio in terminis” (because an irreversibly arrested stem cell ceases to be a stem cell). Senescence is believed to be predominantly induced by activation of p16, and indeed expression of p16 is considered to be a marker for the presence of senescent cells. The genetic or pharmacological depletion of senescent cells in mice has been shown to enhance regenerative potential and extend life span.<sup>57</sup> Although expression of p16 in aged primitive HSC has been contested,<sup>58</sup> pharmacological targeting of senescent bone marrow cells has been shown to have beneficial effects.<sup>59</sup> This suggests that putative senescent cells in the bone marrow may secrete factors that negatively affect HSC potential.

### Polarity

It has been reported that the asymmetric distribution of specific proteins, collectively referred to as “increased polarity,” is a prominent feature of aged HSCs, whereas in young HSCs this is much less obvious.<sup>33,60</sup> Unequal distribution of these proteins is believed to be caused by elevated activity of Cdc42. Interestingly, inhibitors of Cdc42 restore polarity in aged HSCs and improve HSC functioning after transplant. Although it is not straightforward to understand how an acute reversal of the asymmetric distribution of proteins can have long-term effects



**Figure 3. Cell-intrinsic mechanisms that contribute to HSC aging.** Although some of these molecular events are difficult to revert, other may be amenable to pharmacological interventions and could be exploited to rejuvenate HSCs. ROS, reactive oxygen species.

on HSC functioning, this study demonstrates that at least some aspects of the aging process appear to be reversible.

### Impaired autophagy and mitochondrial activity

A large fraction of aged HSCs show impaired levels of autophagy.<sup>33</sup> Autophagy is generally associated with recycling of organelles, and impaired autophagy in aged HSCs appears to specifically result in the accumulation of mitochondria, which in turn induces metabolic stress. It has been demonstrated that high levels of reactive oxygen species, generated by mitochondria, accumulate in aged HSCs and compromise their functioning.<sup>61,62</sup> In addition, reducing mitochondrial stress in aged HSCs can reverse loss of stem functioning.<sup>62</sup> Mitochondrial dysfunctioning caused by accumulating mitochondrial DNA mutations has been shown to cause multiple hematopoietic defects that are typically seen in the elderly, but HSCs themselves appear to be relatively resistant.<sup>63</sup> Taken together, cellular metabolism, controlled by mitochondrial status and reactive oxygen species and mTOR signaling, play an important role in maintaining HSC function throughout life, but the molecular cause of age-dependent metabolic derailment remains unclear. Importantly, however, pharmacological interventions in these signaling pathways are feasible and may be exploited to restore function in aged HSCs.

### Epigenetic reprogramming

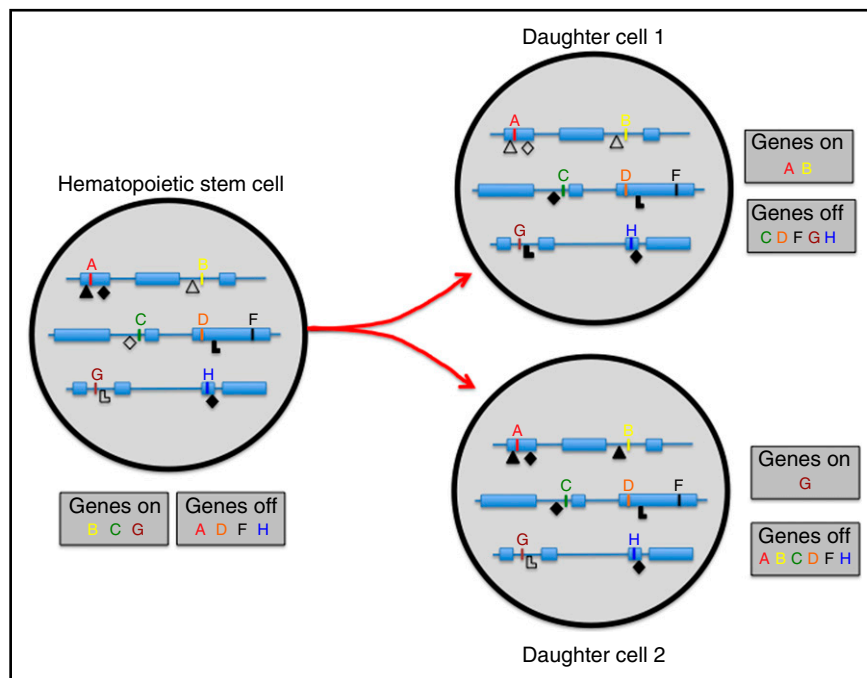
Whereas it is not immediately evident how large-scale random DNA damage would accumulate in aged HSCs, which do barely proliferate, it is not difficult to see how an accumulation of aberrant epigenetic marks could readily but gradually lead to loss of stem cell potential. For a true self-renewal division to occur, an HSC must, in the timeframe of a single division, not only copy and distribute its entire genome across 2 daughter cells, but also the plethora of epigenetic marks that cover each and genomic locus must be faithfully reproduced in at least 1 of the 2 daughter cells. It seems highly likely that not all epigenetic moieties that are required to specify stem cell functioning are properly maintained after a stem cell has divided (Figure 4).

Whereas loss or gain of specific epigenetic marks at defined loci may remain inconsequential, collectively chromatin modifications are important to maintain transcriptional fidelity. The relevance of these epigenetic marks is best exemplified by the fact that perturbation of a large number of epigenetic writers or erasers severely affects HSC function.<sup>64-68</sup> The gradual and random erosion of epigenetic marks as stem cells divide and age, provides a conceptual framework which can explain why, at least in the mouse, stem cells can make only a very limited number of true self-renewal divisions. Because the erosion of epigenetic marks is likely to be different for each stem cell, with age increased functional cell-to-cell variability is expected to develop. Indeed, this is exactly what has been observed in experimental studies.<sup>10,32</sup> Such increased functional heterogeneity is likely the result of increased transcriptional "noise," caused by aberrant epigenetic fidelity. This scenario would also suggest that there are no specific "aging" genes, but rather that aging is caused by the cumulative and combinatorial effect of large collections of stochastically differentially expressed genes. In agreement, several distinct gene expression profile studies have not been able to identify many commonly differentially expressed transcripts, and conversely, transcripts that have shown to be affected during aging in 1 study have often not been confirmed in others.<sup>12,69-73</sup>

The involvement of epigenetic regulation in maintaining proper stem cell transcriptional activity and functioning during aging is also suggested by an increasing number of clinical studies in which mutations in genes encoding for epigenetic enzymes have been found in either elderly people whose hematopoietic system has become oligoclonal, or in patients with myelodysplastic syndromes or acute myeloid leukemia.<sup>52</sup> Epigenetic genes that are relatively frequently affected in these conditions include DNMT3A, EZH2, TET2, and SETDB1.<sup>74-77</sup> Although the enzymatic activity of these proteins is rather well understood (DNA methylation, H3K27 tri-methylation, DNA demethylation, and H3K9 tri-methylation, respectively), their effect on stem cell functioning is far from clear. Without going into depth here on the specific molecular pathways in which these epigenetic writers are involved, it appears very likely that mutations in these



**Figure 4. Repressive (closed symbols) and activating (open symbols) epigenetic marks cover all genes and affect transcriptional status.** If an HSC divides, all genomic and epigenomic information must be properly propagated to daughter cells. If epigenetic marks are lost or gained, genes that should be expressed in HSC (B, C, and G) may become repressed; conversely, genes that should be repressed (A, D, F, and H) may become activated. As a consequence, functional stem cell activity of daughter cell 1 and 2 may be reduced compared with the HSC from which they derive.



genes subtly alter the epigenetic memory of stem cells that, because of large-scale (but again, potentially subtle) transcriptional consequences, increases self-renewal potential of mutant cells. Thus, with time, in the bone marrow cells in which these self-renewal-enhancing mutations have occurred are expected to expand clonally. Indeed, studies in mice have revealed that (enforced) altered expression of wild-type or mutant epigenetic writers affects self-renewal of HSCs.<sup>64,68,78</sup>

## Clonal hematopoiesis in human

As referred to previously, clonal hematopoiesis is a frequent event in elderly people. The first studies that reported on this phenomenon assessed whether X-chromosome inactivation patterns were random or skewed in peripheral blood cells of elderly females.<sup>79,80</sup> Indeed, these early studies suggested that during aging, blood cells are derived from fewer and fewer stem cells. Much more recently, these early findings have been confirmed and significantly extended by multiple independent large-scale sequencing studies.<sup>74-77</sup> In addition, these latter studies have shown that clonal hematopoiesis in otherwise healthy individuals increases the risk to develop leukemia, and, interestingly, cardiovascular disease, and thus is associated with increased mortality. This raises the question of whether clonal hematopoiesis is detrimental, and if so, why? It is important to realize that in many perfectly healthy very aged individuals, prominent clonal hematopoiesis is present without any signs of disease.<sup>74,81</sup> Yet, that age-dependent clonal hematopoiesis is associated with disease suggests that, at least in these individuals, there may be a causal relationship. Whether mutations in epigenetic genes believed to cause clonal hematopoiesis are truly oncogenic remains unclear. It is possible that these mutations merely increase self-renewal of benign HSC and thus lead to clonal expansion of healthy stem cells. Mutations that are truly leukemogenic are then more likely to occur in such a pool of actively self-renewing stem cells; thus, when patients present

with full-blown disease, leukemic cells frequently display mutations in epigenetic genes. In such a scenario, it would probably not be appropriate to refer to these genes as leukemia-initiating events.

That subjects with clonal hematopoiesis are more susceptible to develop nonhematological, most notably cardiovascular, disease, suggests that these mutations also affect the functioning of fully differentiated cells, such as monocytes and macrophages. These end-stage cells have been long known to play a role in vascular remodeling, and an age-dependent impaired functioning could explain their association with cardiovascular problems. In fact, whereas ample attention has focused on the aging of HSC themselves (including the present manuscript), we know much less about the functioning of fully mature peripheral blood cells that are derived from these aged stem cells. It is well established that aged red cells and aged platelets display impaired functioning,<sup>82</sup> but is it also true that red cells and platelets derived from aged stem cells show loss of functioning? It is intriguing that clonal hematopoiesis has been associated with increased incidence of atherosclerotic cardiovascular disease, suggesting that the function of monocytic cells derived from these aged stem cell clones is impaired.<sup>83,84</sup>

Although it has now been well established that clonal hematopoiesis is relatively commonly seen in elderly individuals, it is important to realize that our general understanding of clonal stem cell contribution in hematopoiesis is very limited. We do not know at present how many stem cells actively contribute to blood cell formation during life. Various models have been proposed,<sup>85</sup> ranging from clonal succession (at any given time, a few clones are present that become exhausted and are replaced by new clones),<sup>86</sup> clonal stability (many clones stably contribute and do so throughout life),<sup>87</sup> dynamic repetition (many clones contribute but do so at highly distinct efficiencies),<sup>88</sup> or stochastic behavior (clones contribute randomly, their contribution

may vary dramatically with time, they may become extinct or resurface without any apparent pattern).<sup>89</sup> The uncertainty as to how many stem cells contribute to steady-state hematopoiesis during aging results from the fact that historically clonal descent of blood cells has been difficult to trace. For clonal analyses unique heritable markers must be present that discriminate cells from 1 clone from the other.

Whereas large-scale sequencing efforts that detect spontaneous mutations and their allele frequencies in humans provided insight into clonal makeup in humans, much more precise measurements have been made in experimental conditions using a variety of transgenic approaches. Initial approaches aimed to specifically provide insight into clonal stem cell contribution involved ex vivo barcoding of purified HSC with retro- or lentiviral vectors that contain unique DNA tags, which were subsequently transplanted into recipient mice<sup>90-93</sup> (reviewed in Bystrykh et al<sup>85</sup>). Later studies embarked on in vivo DNA barcoding of stem cells, thereby avoiding the transplantation process.<sup>94,95</sup> Most recently, in vivo clonal marking in transgenic mice and fish has been carried out using fluorescent dyes as tags.<sup>96,97</sup> Potentially as a result of the variety of clonal marking and analyses approaches used, the consistency among these studies is limited. Whereas some studies indicate that only a limited number of stem cells robustly contributes to blood cell formation,<sup>14,96</sup> others suggest that hematopoiesis is highly polyclonal.<sup>90,94</sup> Irrespective of how these differences will ultimately be reconciled, it is evident from many experimental studies that mice in which blood cell production is derived from even a single stem cell, are not necessarily prone to develop leukemia.<sup>10,13,32</sup> Thus, oligo- or even monoclonality is in and of itself not a (preclinical) sign of pathology.

## Is stem cell aging similar in different tissues?

In the field of stem cell biology, comparisons between different stem cell-containing tissues are frequently made, with the assumption that at least some general characteristics of stem cells may be conserved across tissues. Although by definition all tissue-specific stem cells are characterized by their self-renewal potential, it is not at all clear whether stem cells age similarly in distinct tissues. In fact, in comparing intestinal stem cells with those of the hematopoietic system, it appears as if aging is very different in these 2 tissues. Whereas in the hematopoietic system, stem cell turnover is very low, proliferation rates in the intestine are very high.<sup>98</sup> Intestinal stem cells do, as expected, accumulate random DNA damage,<sup>99</sup> but they do not show functional decline during normal aging.<sup>100</sup> It will be interesting to assess why rapidly turning over intestinal stem cells do not age and slow-turning over HSC do. In this respect, it is interesting that another important difference between blood and gut is the extent to which plasticity occurs in progenitor cells in these respective tissues. In the intestine, progenitors have been shown to be able to revert to a stem cell fate,<sup>101</sup> but in the hematopoietic system such progenitor-to-stem cell conversions have never been described, at least not during steady-state in vivo hematopoiesis. However, enforced expression of transcription factors or microRNAs has been shown to be able to generate transplantable stem cells from committed cell types.<sup>102-104</sup>

## Future perspectives

Assuming that in human HSC aging is not fundamentally different than in mice, it is interesting to speculate that the age-dependent increase of the stem cell pool size is an important factor in the increase of prevalence of myeloproliferative diseases and leukemia in the elderly. An expansion of the pool of primitive cells would increase the number of target cells for malignant derailment. Alternatively, aged stem cells would intrinsically be more susceptible to malignant transformation, possibly as a result of an altered epigenetic landscape that may have accumulated as a result of repeated proliferation. Also, there is some evidence (but not a lot) that the same oncogenic mutation results in a more aggressive disease in aged stem cells compared with young cells.<sup>105</sup> If indeed, as in mice, aged human stem cells undergo very few divisions, it seems implausible that these primitive cells themselves are prone to accumulate multiple mutations.

Would monitoring of HSC functionality during human aging be possible or indeed useful? At this point, the answer to both of these questions is negative; there is no reliable assay that can quantify human stem cell aging, and if there were such an assay, there are no obvious intervention strategies available. However, our understanding of HSC aging is increasing very rapidly, and we may in the near future well be able to identify individuals who display enhanced stem cell aging. This may offer opportunities to intervene in the kinetics with which HSCs age. Antiaging interventions may be aimed to prevent, delay, or, most speculatively, reverse aspects of the stem cell aging process. Studies in other tissues have demonstrated that at least delaying components of the aging process can be a viable strategy,<sup>106</sup> and there is no fundamental reason to believe that the hematopoietic system would be exempt for such approaches. In fact, reversal of some aging phenotypes has been achieved by reprogramming old HSCs to pluripotency and subsequently generating definitive hematopoiesis from these induced pluripotent stem cells.<sup>107</sup> Future interventions may be dietary, pharmacological, or eventually cell therapeutically.

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## Authorship

Contribution: G.d.H. and S.S.L. wrote the paper.

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## Footnote

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## REFERENCES

1. Ossenkoppele G, Löwenberg B. How I treat the older patient with acute myeloid leukemia. *Blood*. 2015;125(5):767-774.
2. López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell*. 2013;153(6):1194-1217.
3. Harrison DE, Astle CM, Stone M. Numbers and functions of transplantable primitive immunohematopoietic stem cells. Effects of age. *J Immunol*. 1989;142(11):3833-3840.
4. Stiehm ER, Roberts RL, Hanley-Lopez J, et al. Bone marrow transplantation in severe combined immunodeficiency from a sibling who had received a paternal bone marrow transplant. *N Engl J Med*. 1996;335(24):1811-1814.
5. Harrison DE, Astle CM. Loss of stem cell repopulating ability upon transplantation. Effects of donor age, cell number, and transplantation procedure. *J Exp Med*. 1982;156(6):1767-1779.
6. Hellman S, Botnick LE, Hannon EC, Vignoulle RM. Proliferative capacity of murine hematopoietic stem cells. *Proc Natl Acad Sci USA*. 1978;75(1):490-494.
7. Mauch P, Botnick LE, Hannon EC, Obbagy J, Hellman S. Decline in bone marrow proliferative capacity as a function of age. *Blood*. 1982;60(1):245-252.
8. Ema H, Sudo K, Seita J, et al. Quantification of self-renewal capacity in single hematopoietic stem cells from normal and Lnk-deficient mice. *Dev Cell*. 2005;8(6):907-914.
9. Wolf NS, Priestley GV, Averill LE. Depletion of reserve in the hemopoietic system: III. Factors affecting the serial transplantation of bone marrow. *Exp Hematol*. 1983;11(8):762-771.
10. Dykstra B, Olthof S, Schreuder J, Ritsema M, de Haan G. Clonal analysis reveals multiple functional defects of aged murine hematopoietic stem cells. *J Exp Med*. 2011;208(13):2691-2703.
11. Van Zant G, Holland BP, Eldridge PW, Chen JJ. Genotype-restricted growth and aging patterns in hematopoietic stem cell populations of allophenic mice. *J Exp Med*. 1990;171(5):1547-1565.
12. Rossi DJ, Bryder D, Zahn JM, et al. Cell intrinsic alterations underlie hematopoietic stem cell aging. *Proc Natl Acad Sci USA*. 2005;102(26):9194-9199.
13. Sudo K, Ema H, Morita Y, Nakauchi H. Age-associated characteristics of murine hematopoietic stem cells. *J Exp Med*. 2000;192(9):1273-1280.
14. Verovskaya E, Broekhuis MJ, Zwart E, et al. Heterogeneity of young and aged murine hematopoietic stem cells revealed by quantitative clonal analysis using cellular barcoding. *Blood*. 2013;122(4):523-532.
15. Morrison SJ, Wandycz AM, Akashi K, Globerson A, Weissman IL. The aging of hematopoietic stem cells. *Nat Med*. 1996;2(9):1011-1016.
16. Geiger H, True JM, de Haan G, Van Zant G. Age- and stage-specific regulation patterns in the hematopoietic stem cell hierarchy. *Blood*. 2001;98(10):2966-2972.
17. Beerman I, Bhattacharya D, Zandi S, et al. Functionally distinct hematopoietic stem cells modulate hematopoietic lineage potential during aging by a mechanism of clonal expansion. *Proc Natl Acad Sci USA*. 2010;107(12):5465-5470.
18. Van Zant G. Studies of hematopoietic stem cells spared by 5-fluorouracil. *J Exp Med*. 1984;159(3):679-690.
19. Bradford GB, Williams B, Rossi R, Bertoncello I. Quiescence, cycling, and turnover in the primitive hematopoietic stem cell compartment. *Exp Hematol*. 1997;25(5):445-453.
20. Cheshier SH, Morrison SJ, Liao X, Weissman IL. In vivo proliferation and cell cycle kinetics of long-term self-renewing hematopoietic stem cells. *Proc Natl Acad Sci USA*. 1999;96(6):3120-3125.
21. Dykstra B, Kent D, Bowie M, et al. Long-term propagation of distinct hematopoietic differentiation programs in vivo. *Cell Stem Cell*. 2007;1(2):218-229.
22. Morita Y, Ema H, Nakauchi H. Heterogeneity and hierarchy within the most primitive hematopoietic stem cell compartment. *J Exp Med*. 2010;207(6):1173-1182.
23. Bernitz JM, Kim HS, MacArthur B, Sieburg H, Moore K. Hematopoietic stem cells count and remember self-renewal divisions. *Cell*. 2016;167(5):1296-1309.
24. Van Zant G, de Haan G, Rich IN. Alternatives to stem cell renewal from a developmental viewpoint. *Exp Hematol*. 1997;25(3):187-192.
25. Jung JJ, Buisman SC, de Haan G. Do hematopoietic stem cells get old? *Leukemia*. 2017;31(3):529-531.
26. Vaziri H, Dragowska W, Allsopp RC, Thomas TE, Harley CB, Lansdorf PM. Evidence for a mitotic clock in human hematopoietic stem cells: loss of telomeric DNA with age. *Proc Natl Acad Sci USA*. 1994;91(21):9857-9860.
27. Kuranda K, Vargaftig J, de la Rochere P, et al. Age-related changes in human hematopoietic stem/progenitor cells. *Aging Cell*. 2011;10(3):542-546.
28. Pang WW, Price EA, Sahoo D, et al. Human bone marrow hematopoietic stem cells are increased in frequency and myeloid-biased with age. *Proc Natl Acad Sci USA*. 2011;108(50):20012-20017.
29. Rundberg Nilsson A, Soneji S, Adolfsson S, Bryder D, Pronk CJ. Human and murine hematopoietic stem cell aging is associated with functional impairments and intrinsic megakaryocytic/erythroid bias. *PLoS One*. 2016;11(7):e0158369.
30. De Haan G, Van Zant G. Genetic analysis of hemopoietic cell cycling in mice suggests its involvement in organismal life span. *FASEB J*. 1999;13(6):707-713.
31. de Haan G, Van Zant G. Intrinsic and extrinsic control of hemopoietic stem cell numbers: mapping of a stem cell gene. *J Exp Med*. 1997;186(4):529-536.
32. Cho RH, Sieburg HB, Muller-Sieburg CE. A new mechanism for the aging of hematopoietic stem cells: aging changes the clonal composition of the stem cell compartment but not individual stem cells. *Blood*. 2008;111(12):5553-5561.
33. Ho TT, Warr MR, Adelman ER, et al. Autophagy maintains the metabolism and function of young and old stem cells. *Nature*. 2017;543(7644):205-210.
34. Wilson NK, Kent DG, Buettner F, et al. Combined single-cell functional and gene expression analysis resolves heterogeneity within stem cell populations. *Cell Stem Cell*. 2015;16(6):712-724.
35. Grover A, Sanjuan-Pla A, Thongjuea S, et al. Single-cell RNA sequencing reveals molecular and functional platelet bias of aged haematopoietic stem cells. *Nat Commun*. 2016;7:11075.
36. Kowalczyk MS, Tirosh I, Heckl D, et al. Single-cell RNA-seq reveals changes in cell cycle and differentiation programs upon aging of hematopoietic stem cells. *Genome Res*. 2015;25(12):1860-1872.
37. Mayack SR, Shadrach JL, Kim FS, Wagers AJ. Systemic signals regulate ageing and rejuvenation of blood stem cell niches [article retracted in *Nature*. 2010;467(7317):872]. *Nature*. 2010;463(7280):495-500.
38. Katsimpardi L, Litterman NK, Schein PA, et al. Vascular and neurogenic rejuvenation of the aging mouse brain by young systemic factors. *Science*. 2014;344(6184):630-634.
39. Conboy MJ, Conboy IM, Rando TA. Heterochronic parabiosis: historical perspective and methodological considerations for studies of aging and longevity. *Aging Cell*. 2013;12(3):525-530.
40. Villeda SA, Plambeck KE, Middeldorp J, et al. Young blood reverses age-related impairments in cognitive function and synaptic plasticity in mice. *Nat Med*. 2014;20(6):659-663.
41. Castellano JM, Mosher KI, Abbey RJ, et al. Human umbilical cord plasma proteins revitalize hippocampal function in aged mice. *Nature*. 2017;544(7651):488-492.
42. Kovtonyuk LV, Fritsch K, Feng X, Manz MG, Takizawa H. Inflamm-aging of hematopoiesis, hematopoietic stem cells, and the bone marrow microenvironment. *Front Immunol*. 2016;7:502.
43. Ergen AV, Boles NC, Goodell MA. Rantes/Ccl5 influences hematopoietic stem cell subtypes and causes myeloid skewing. *Blood*. 2012;119(11):2500-2509.
44. Zambetti NA, Ping Z, Chen S, et al. Mesenchymal inflammation drives genotoxic stress in hematopoietic stem cells and predicts disease evolution in human pre-leukemia. *Cell Stem Cell*. 2016;19(5):613-627.
45. Geiger H, de Haan G, Florian MC. The ageing haematopoietic stem cell compartment. *Nat Rev Immunol*. 2013;13(5):376-389.
46. Ju Z, Jiang H, Jaworski M, et al. Telomere dysfunction induces environmental alterations limiting hematopoietic stem cell function and engraftment. *Nat Med*. 2007;13(6):742-747.

47. Rossi DJ, Bryder D, Seita J, Nussenzweig A, Hoeijmakers J, Weissman IL. Deficiencies in DNA damage repair limit the function of haematopoietic stem cells with age. *Nature*. 2007;447(7145):725-729.
48. Alter BP, Rosenberg PS, Giri N, Baerlocher GM, Lansdorp PM, Savage SA. Telomere length is associated with disease severity and declines with age in dyskeratosis congenita. *Haematologica*. 2012;97(3):353-359.
49. Walter D, Lier A, Geiselhart A, et al. Exit from dormancy provokes DNA-damage-induced attrition in haematopoietic stem cells. *Nature*. 2015;520(7548):549-552.
50. Yamamoto R, Morita Y, Oeohara J, et al. Clonal analysis unveils self-renewing lineage-restricted progenitors generated directly from hematopoietic stem cells. *Cell*. 2013;154(5):1112-1126.
51. Flach J, Bakker ST, Mohrin M, et al. Replication stress is a potent driver of functional decline in ageing haematopoietic stem cells. *Nature*. 2014;512(7513):198-202.
52. Steensma DP, Bejar R, Jaiswal S, et al. Clonal hematopoiesis of indeterminate potential and its distinction from myelodysplastic syndromes. *Blood*. 2015;126(1):9-16.
53. Aubert G, Lansdorp PM. Telomeres and aging. *Physiol Rev*. 2008;88(2):557-579.
54. Rudolph KL, Chang S, Lee HW, et al. Longevity, stress response, and cancer in aging telomerase-deficient mice. *Cell*. 1999;96(5):701-712.
55. Allsopp RC, Morin GB, Horner JW, DePinho R, Harley CB, Weissman IL. Effect of TERT over-expression on the long-term transplantation capacity of hematopoietic stem cells. *Nat Med*. 2003;9(4):369-371.
56. Childs BG, Durik M, Baker DJ, van Deursen JM. Cellular senescence in aging and age-related disease: from mechanisms to therapy. *Nat Med*. 2015;21(12):1424-1435.
57. Baker DJ, Childs BG, Durik M, et al. Naturally occurring p16(Ink4a)-positive cells shorten healthy lifespan. *Nature*. 2016;530(7589):184-189.
58. Attema JL, Pronk CJ, Norddahl GL, Nygren JM, Bryder D. Hematopoietic stem cell ageing is uncoupled from p16 INK4A-mediated senescence. *Oncogene*. 2009;28(22):2238-2243.
59. Chang J, Wang Y, Shao L, et al. Clearance of senescent cells by ABT263 rejuvenates aged hematopoietic stem cells in mice. *Nat Med*. 2016;22(1):78-83.
60. Florian MC, Dörr K, Niebel A, et al. Cdc42 activity regulates hematopoietic stem cell aging and rejuvenation. *Cell Stem Cell*. 2012;10(5):520-530.
61. Ito K, Hirao A, Arai F, et al. Reactive oxygen species act through p38 MAPK to limit the lifespan of hematopoietic stem cells. *Nat Med*. 2006;12(4):446-451.
62. Mohrin M, Shin J, Liu Y, et al. Stem cell aging. A mitochondrial UPR-mediated metabolic checkpoint regulates hematopoietic stem cell aging. *Science*. 2015;347(6228):1374-1377.
63. Norddahl GL, Pronk CJ, Wahlestedt M, et al. Accumulating mitochondrial DNA mutations drive premature hematopoietic aging phenotypes distinct from physiological stem cell aging. *Cell Stem Cell*. 2011;8(5):499-510.
64. Kamminga LM, Bystrykh LV, de Boer A, et al. The polycomb group gene Ezh2 prevents hematopoietic stem cell exhaustion. *Blood*. 2006;107(5):2170-2179.
65. Iwama A, Oguro H, Negishi M, et al. Enhanced self-renewal of hematopoietic stem cells mediated by the polycomb gene product Bmi-1. *Immunity*. 2004;21(6):843-851.
66. Lessard J, Schumacher A, Thorsteinsdottir U, van Lohuizen M, Magnuson T, Sauvageau G. Functional antagonism of the polycomb-group genes eed and Bmi1 in hematopoietic cell proliferation. *Genes Dev*. 1999;13(20):2691-2703.
67. Majewski IJ, Ritchie ME, Phipson B, et al. Opposing roles of polycomb repressive complexes in hematopoietic stem and progenitor cells. *Blood*. 2010;116(5):731-739.
68. Klauke K, Radulović V, Broekhuis M, et al. Polycomb Cbx family members mediate the balance between haematopoietic stem cell self-renewal and differentiation. *Nat Cell Biol*. 2013;15(4):353-362.
69. Noda S, Ichikawa H, Miyoshi H. Hematopoietic stem cell aging is associated with functional decline and delayed cell cycle progression. *Biochem Biophys Res Commun*. 2009;383(2):210-215.
70. Wahlestedt M, Norddahl GL, Sten G, et al. An epigenetic component of hematopoietic stem cell aging amenable to reprogramming into a young state. *Blood*. 2013;121(21):4257-4264.
71. Beerman I, Bock C, Garrison BS, et al. Proliferation-dependent alterations of the DNA methylation landscape underlie hematopoietic stem cell aging. *Cell Stem Cell*. 2013;12(4):413-425.
72. Sun D, Luo M, Jeong M, et al. Epigenomic profiling of young and aged HSCs reveals concerted changes during aging that reinforce self-renewal. *Cell Stem Cell*. 2014;14(5):673-688.
73. Chambers SM, Shaw CA, Gatza C, Fisk CJ, Donehower LA, Goodell MA. Aging hematopoietic stem cells decline in function and exhibit epigenetic dysregulation. *PLoS Biol*. 2007;5(8):e201.
74. Zink F, Stacey SN, Norddahl GL, et al. Clonal hematopoiesis, with and without candidate driver mutations, is common in the elderly. *Blood*. 2017;130(6):742-752.
75. Xie M, Lu C, Wang J, et al. Age-related mutations associated with clonal hematopoietic expansion and malignancies. *Nat Med*. 2014;20(12):1472-1478.
76. Jaiswal S, Fontanillas P, Flannick J, et al. Age-related clonal hematopoiesis associated with adverse outcomes. *N Engl J Med*. 2014;371(26):2488-2498.
77. Genovese G, Kähler AK, Handsaker RE, et al. Clonal hematopoiesis and blood-cancer risk inferred from blood DNA sequence. *N Engl J Med*. 2014;371(26):2477-2487.
78. Mayle A, Yang L, Rodriguez B, et al. Dnmt3a loss predisposes murine hematopoietic stem cells to malignant transformation. *Blood*. 2015;125(4):629-638.
79. Busque L, Mio R, Mattioli J, et al. Nonrandom X-inactivation patterns in normal females: lyonization ratios vary with age. *Blood*. 1996;88(1):59-65.
80. Champion KM, Gilbert JG, Asimakopoulos FA, Hinshelwood S, Green AR. Clonal haemopoiesis in normal elderly women: implications for the myeloproliferative disorders and myelodysplastic syndromes. *Br J Haematol*. 1997;97(4):920-926.
81. van den Akker EB, Pitts SJ, Deelen J, et al.; Genome of The Netherlands Consortium. Uncompromised 10-year survival of oldest old carrying somatic mutations in DNMT3A and TET2. *Blood*. 2016;127(11):1512-1515.
82. Hoehn RS, Jernigan PL, Chang AL, Edwards MJ, Pritts TA. Molecular mechanisms of erythrocyte aging. *Biol Chem*. 2015;396(6-7):621-631.
83. Fuster JJ, MacLauchlan S, Zuriaga MA, et al. Clonal hematopoiesis associated with TET2 deficiency accelerates atherosclerosis development in mice. *Science*. 2017;355(6327):842-847.
84. Jaiswal S, Natarajan P, Silver AJ, et al. Clonal Hematopoiesis and Risk of Atherosclerotic Cardiovascular Disease. *N Engl J Med*. 2017;377(2):111-121.
85. Bystrykh LV, Verovskaya E, Zwart E, Broekhuis M, de Haan G. Counting stem cells: methodological constraints. *Nat Methods*. 2012;9(6):567-574.
86. Rosendaal M, Adam J. Haemopoiesis by clonal succession? *Blood Cells*. 1984;10(2-3):473-485.
87. Prchal JT, Prchal JF, Belickova M, et al. Clonal stability of blood cell lineages indicated by X-chromosomal transcriptional polymorphism. *J Exp Med*. 1996;183(2):561-567.
88. Takizawa H, Regoes RR, Boddupalli CS, Bonhoeffer S, Manz MG. Dynamic variation in cycling of hematopoietic stem cells in steady state and inflammation. *J Exp Med*. 2011;208(2):273-284.
89. Roeder I, Kamminga LM, Braesels K, Dontje B, de Haan G, Loeffler M. Competitive clonal hematopoiesis in mouse chimeras explained by a stochastic model of stem cell organization. *Blood*. 2005;105(2):609-616.
90. Lu R, Neff NF, Quake SR, Weissman IL. Tracking single hematopoietic stem cells in vivo using high-throughput sequencing in conjunction with viral genetic barcoding. *Nat Biotechnol*. 2011;29(10):928-933.
91. Gerrits A, Dykstra B, Kalmykova OJ, et al. Cellular barcoding tool for clonal analysis in the hematopoietic system. *Blood*. 2010;115(13):2610-2618.
92. Naik SH, Perié L, Swart E, et al. Diverse and heritable lineage imprinting of early haematopoietic progenitors. *Nature*. 2013;496(7444):229-232.
93. Cheung AM, Nguyen LV, Carles A, et al. Analysis of the clonal growth and



- p>differentiation dynamics of primitive bar-coded human cord blood cells in NSG mice.
- Blood*
- . 2013;122(18):3129-3137.
94. Sun J, Ramos A, Chapman B, et al. Clonal dynamics of native haematopoiesis. *Nature*. 2014;514(7522):322-327.
95. Busch K, Klapproth K, Barile M, et al. Fundamental properties of unperturbed haematopoiesis from stem cells in vivo. *Nature*. 2015;518(7540):542-546.
96. Yu VW, Yusuf RZ, Oki T, et al. Epigenetic memory underlies cell-autonomous heterogeneous behavior of hematopoietic stem cells [published correction appears in *Cell*. 2017;168(5):944-945]. *Cell*. 2016;167(5):1310-1322.
97. Henninger J, Santoso B, Hans S, et al. Clonal fate mapping quantifies the number of haematopoietic stem cells that arise during development. *Nat Cell Biol*. 2017;19(1):17-27.
98. Clevers H. The intestinal crypt, a prototype stem cell compartment. *Cell*. 2013;154(2):274-284.
99. Blokzijl F, de Ligt J, Jager M, et al. Tissue-specific mutation accumulation in human adult stem cells during life. *Nature*. 2016;538(7624):260-264.
100. Barker N, Huch M, Kujala P, et al. Lgr5(+ve) stem cells drive self-renewal in the stomach and build long-lived gastric units in vitro. *Cell Stem Cell*. 2010;6(1):25-36.
101. Tetteh PW, Basak O, Farin HF, et al. Replacement of lost Lgr5-positive stem cells through plasticity of their enterocyte-lineage daughters. *Cell Stem Cell*. 2016;18(2):203-213.
102. Riddell J, Gazit R, Garrison BS, et al. Reprogramming committed murine blood cells to induced hematopoietic stem cells with defined factors [published correction appears in *Cell*. 2014;158(1):226]. *Cell*. 2014;157(3):549-564.
103. Pereira CF, Chang B, Qiu J, et al. Induction of a hemogenic program in mouse fibroblasts. *Cell Stem Cell*. 2013;13(2):205-218.
104. Wojtowicz EE, Lechman ER, Hermans KG, et al. Ectopic miR-125a expression induces long-term repopulating stem cell capacity in mouse and human hematopoietic progenitors. *Cell Stem Cell*. 2016;19(3):383-396.
105. Signer RA, Montecino-Rodriguez E, Witte ON, McLaughlin J, Dorshkind K. Age-related defects in B lymphopoiesis underlie the myeloid dominance of adult leukemia. *Blood*. 2007;110(6):1831-1839.
106. Goodell MA, Rando TA. Stem cells and healthy aging. *Science*. 2015;350(6265):1199-1204.
107. Wahlestedt M, Erlandsson E, Kristiansen T, et al. Clonal reversal of ageing-associated stem cell lineage bias via a pluripotent intermediate. *Nat Commun*. 2017;8:14533.



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